

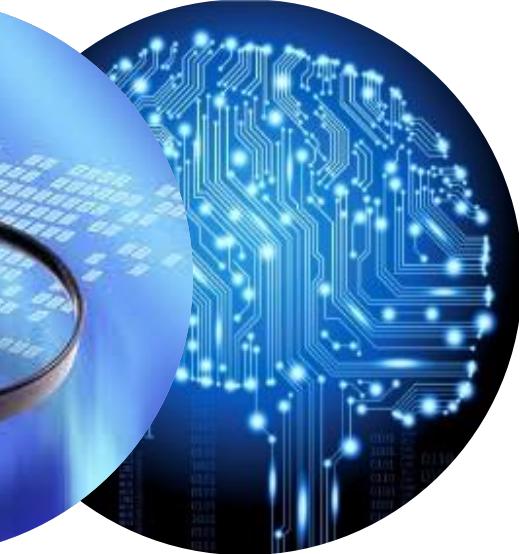
# Safer Food with BIG data

*The Bigger, the Better?*



*Alex Bossers (WBVR) & Martijn Bouwknegt (Vion)*

1H4F Najaarscongres  
8 November 2018

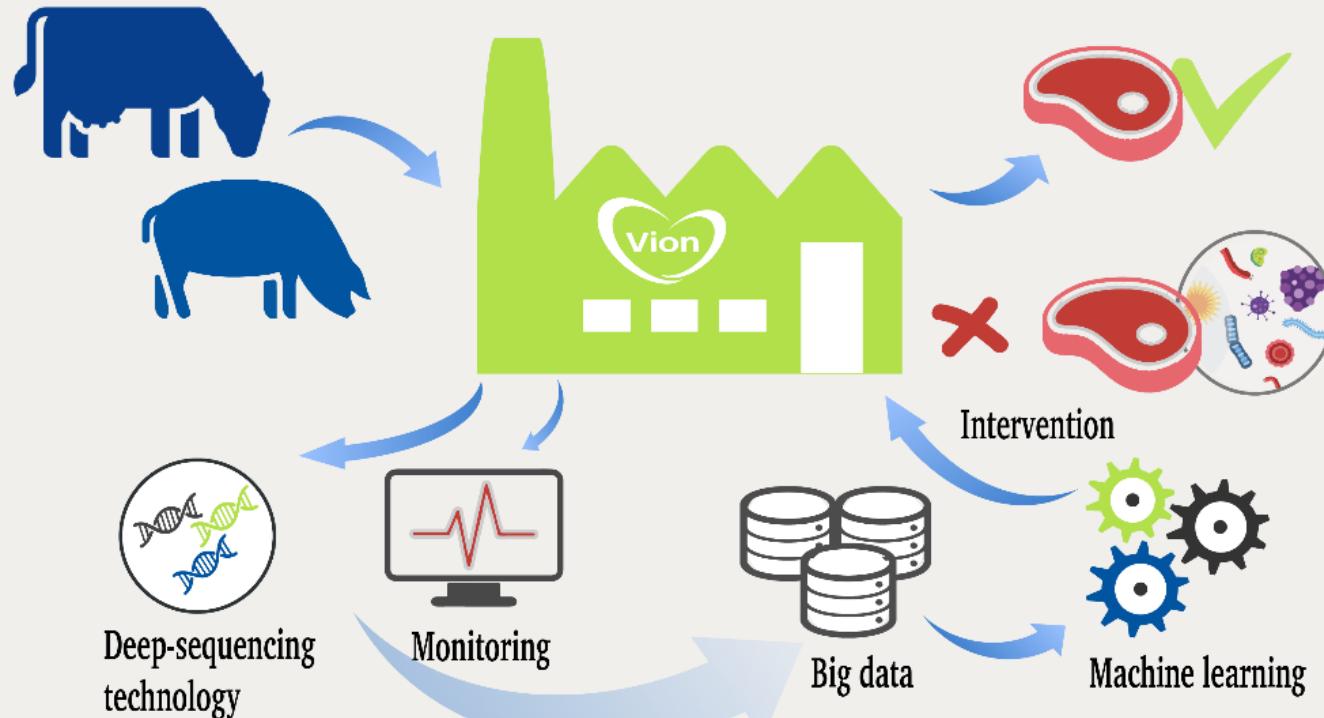


# Summary



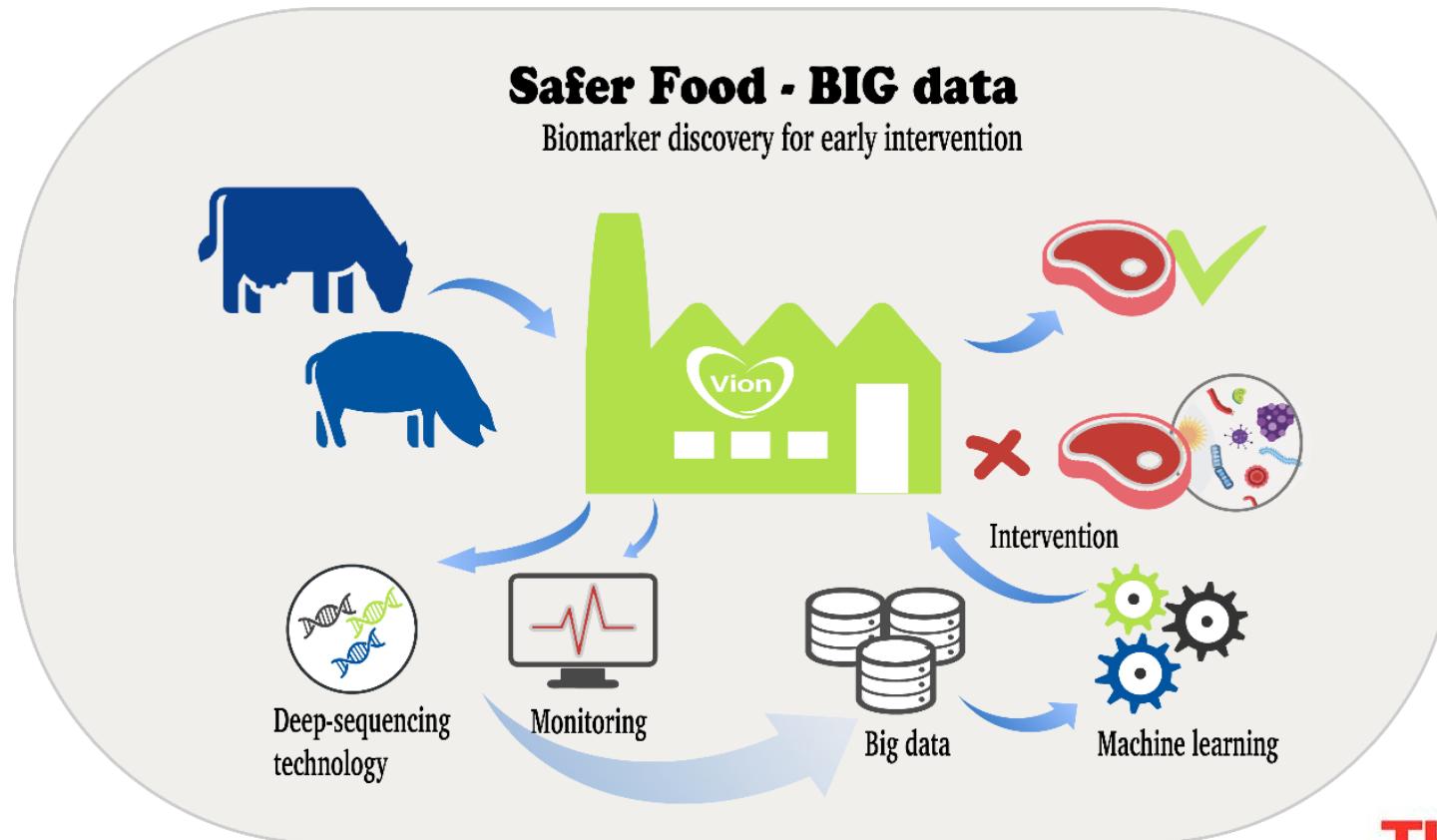
## Safer Food - BIG data

Biomarker discovery for early intervention



- Evaluation of methodologies for sample processing
- Deep sequencing and biomarker discovery
- Pathogen clonality analysis by WGS
- WBVR link WU Food Microbiology laboratory

- Data storage and integration
- WATSON: Machine learning - Identify factors leading to emergence of food-born pathogens



- Delivery of samples,
- meta-data on environmental, physical, biological and chemical variables.
- Intervention strategies
- ...

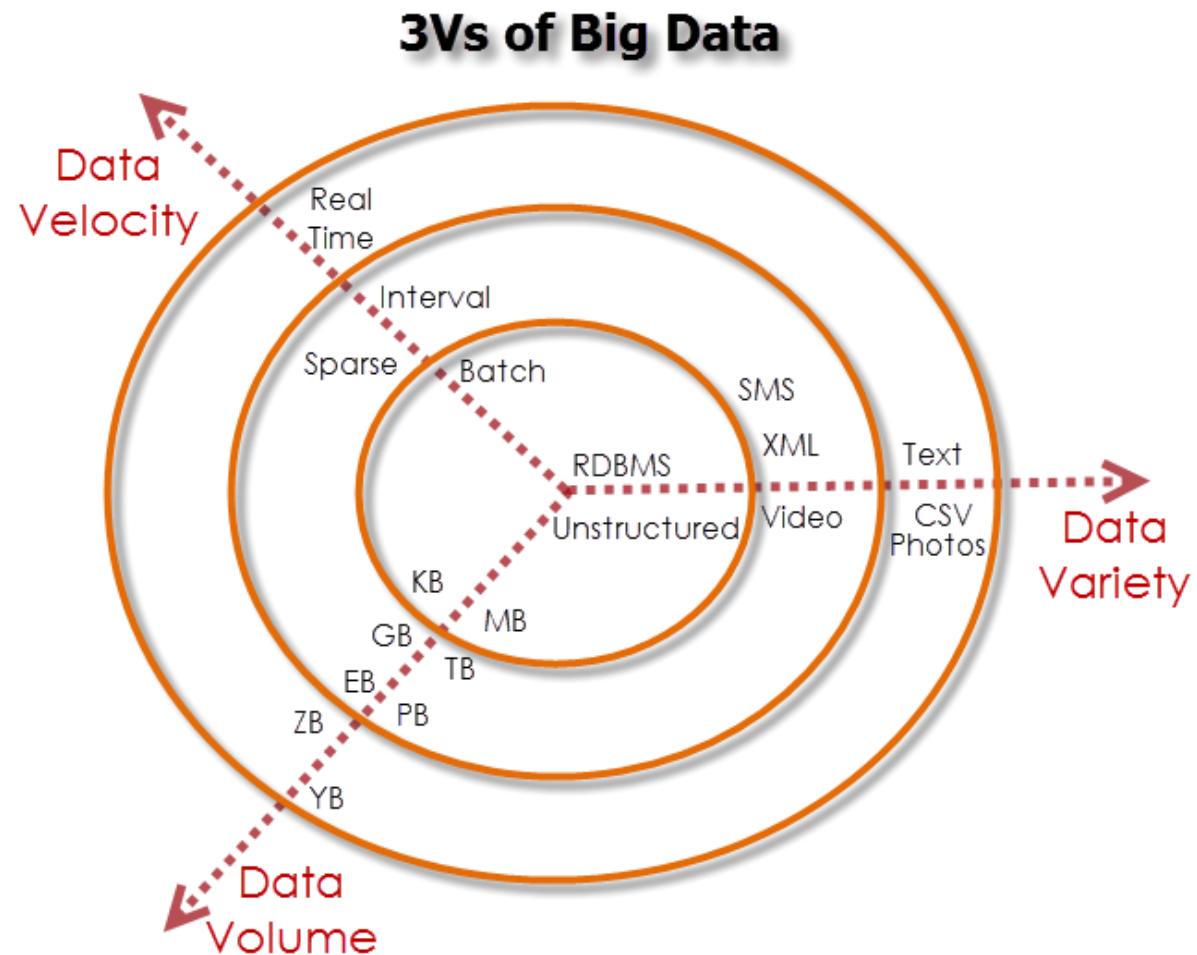
- Knowledge and tools for sample preservation, processing and preparation of RNA/DNA
- Biomarker screening / detection kits, sample prep kits.

# BIG data



## ■ 3Vs

- Velocity
- Volume
- Variety



# BIG data



## ■ 3Vs

- Velocity
- Volume
- Variety

## ■ Types

- Sensor data (temperature, humidity, belt speed)
- Continuous generic microbial monitoring
- Specific data types like microbiomes, imaging, ...



# PPS – Safer Food with BIG data



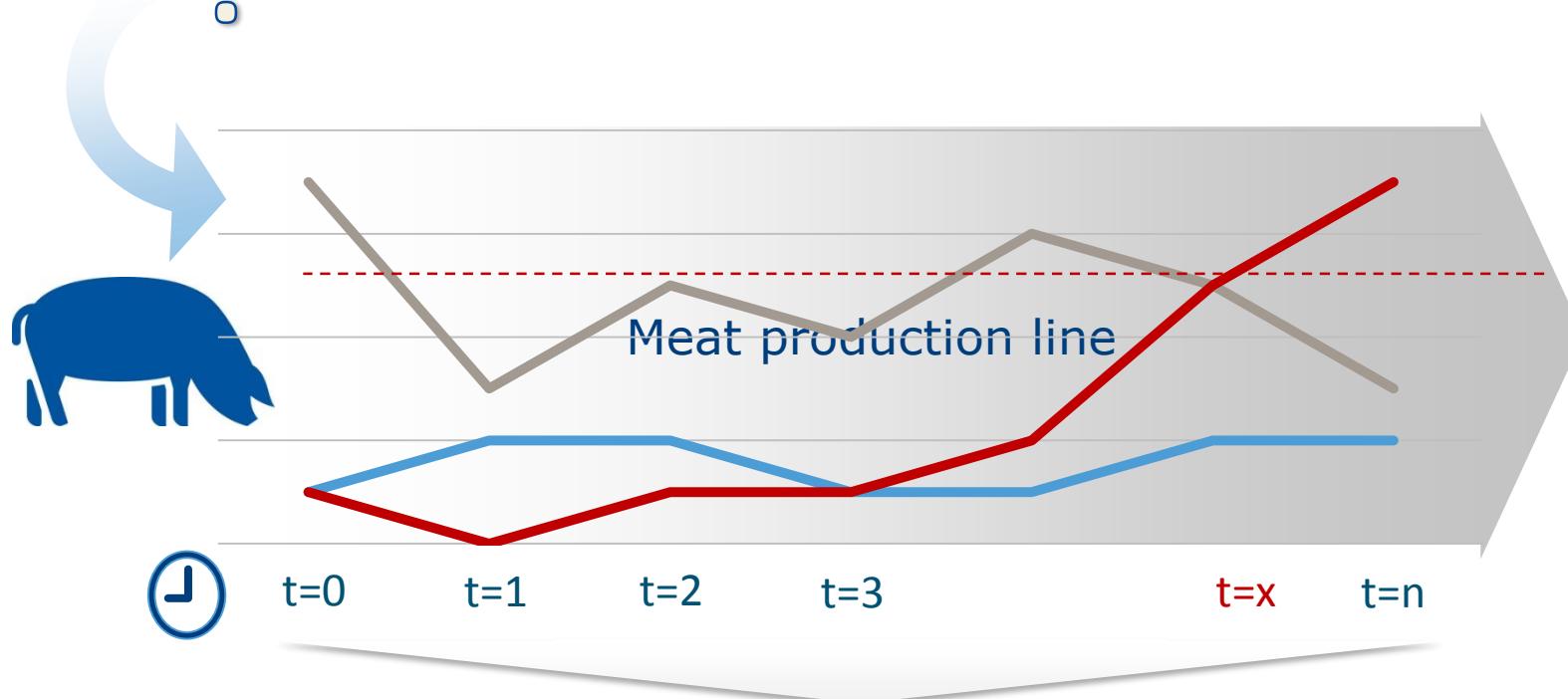
- Develop blue print how to use BIG data approaches to enable early detection of future potential product contamination
  - Blue print can be re-used for other agents / species / products / slaughterhouses
- Demonstration use case:
  - Prevention of Salmonella contamination pig meat
  - Develop methods to determine early (bio)markers
    - Using BIG data methodologies
  - Model intervention strategies

## “DATA”

Environmental factors (humidity/temperature)

Other “routine” lab-measurements

Microbiomes

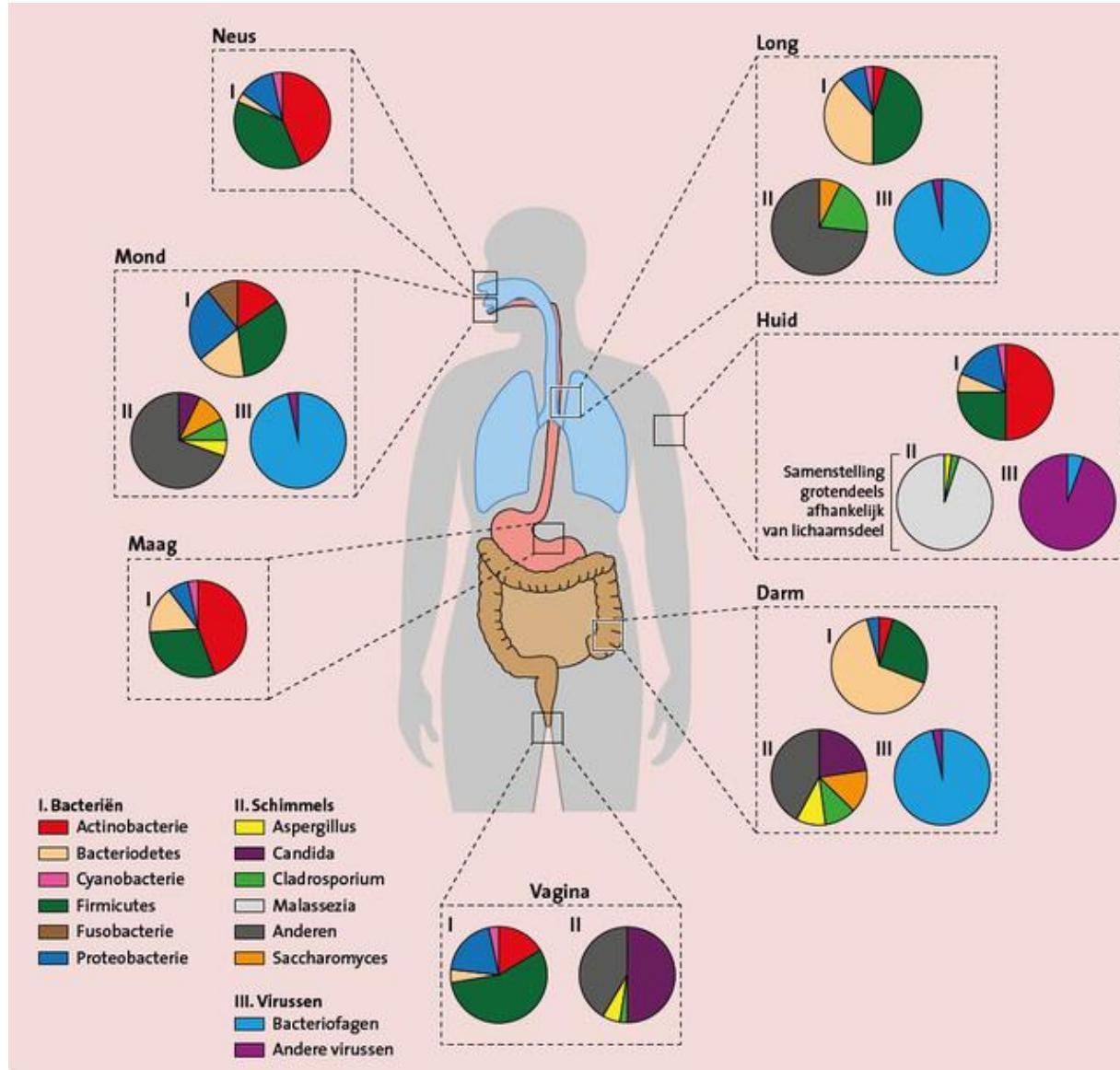


# Microbiome



# Microbioom samenstelling - functie

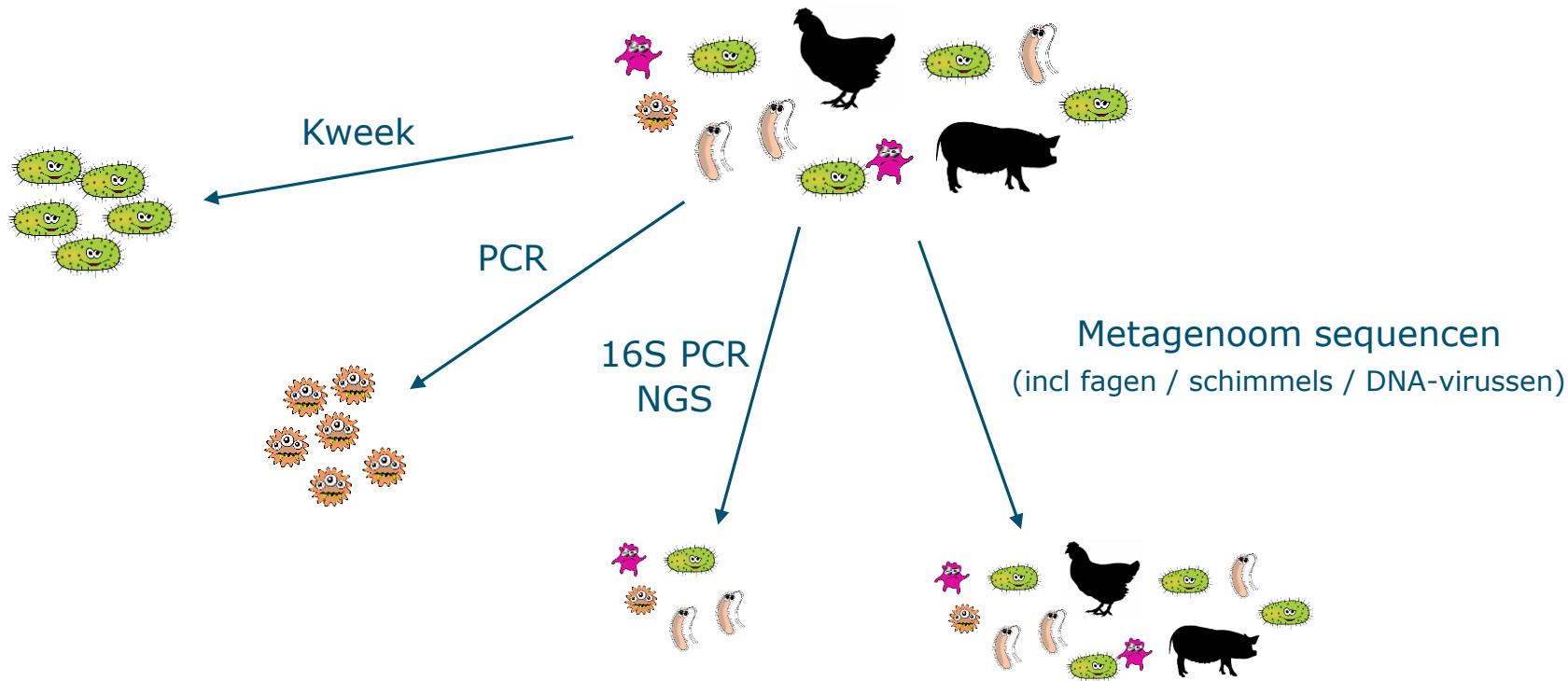
## Spatio- (en temporale) variabiliteit



# Microbioom screenen

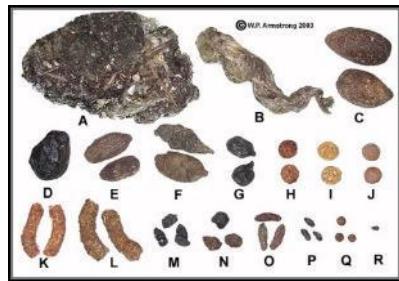


- Metagenoom = alle aanwezige DNA (ook van evt gastheer/omgeving)



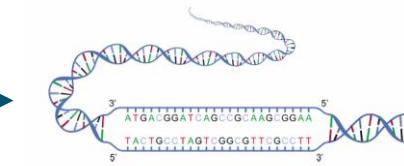
# Metagenomic profiling

## Deep-sequencing



Environmental  
sample

DNA extraction



Total DNA  
(chromosomal + plasmid)

Fragmentation  
Library generation

16S-amplicons



Deep-sequencing

Typical metagenome analysis

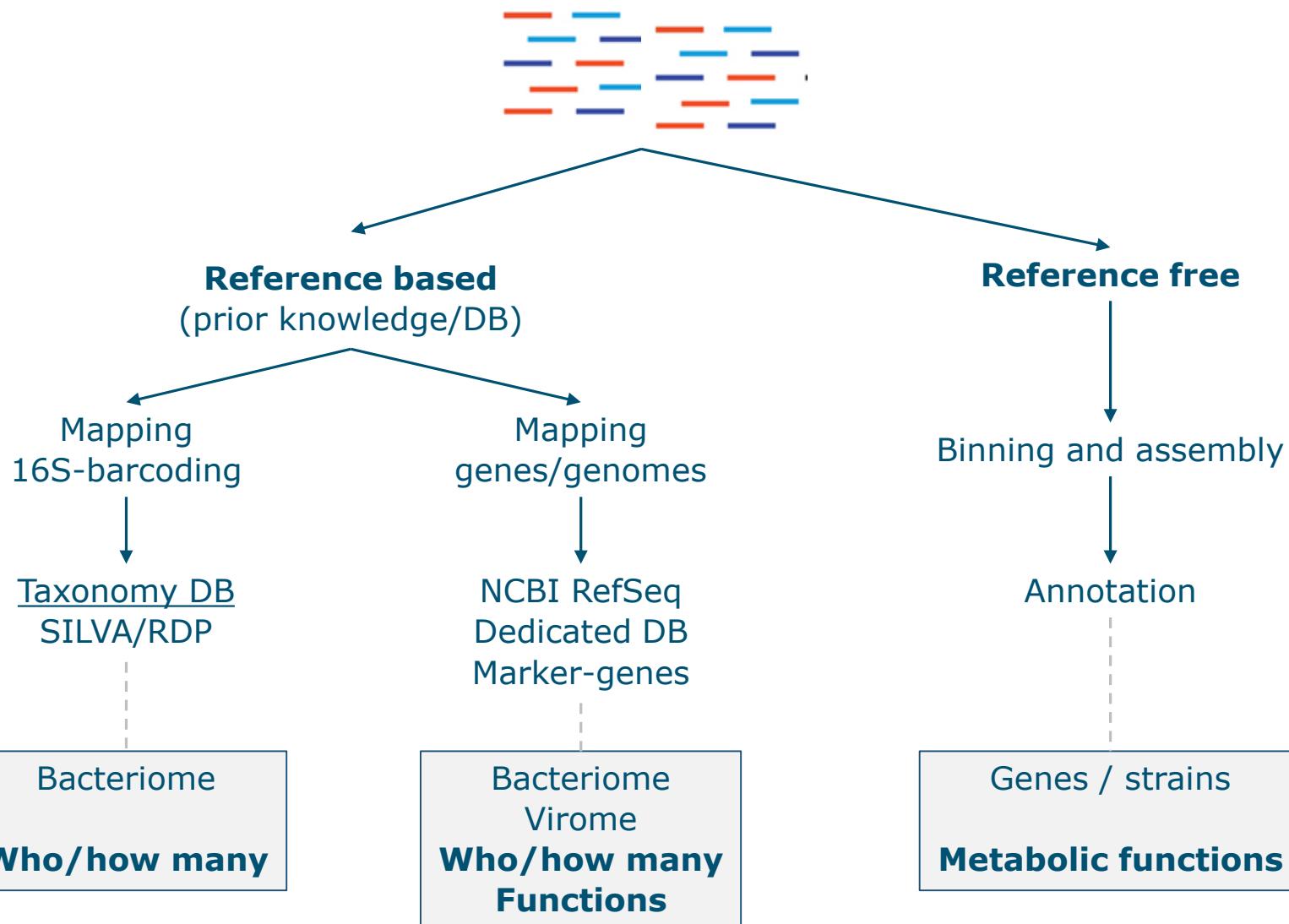
(40M DNA clusters PE150)  
(12 billion bases per sample)

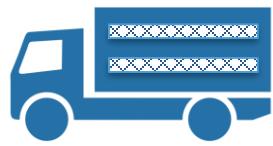
**~24 billion data points per sample**

**Millions of DNA  
reads**  
**~ 150-300 bp**

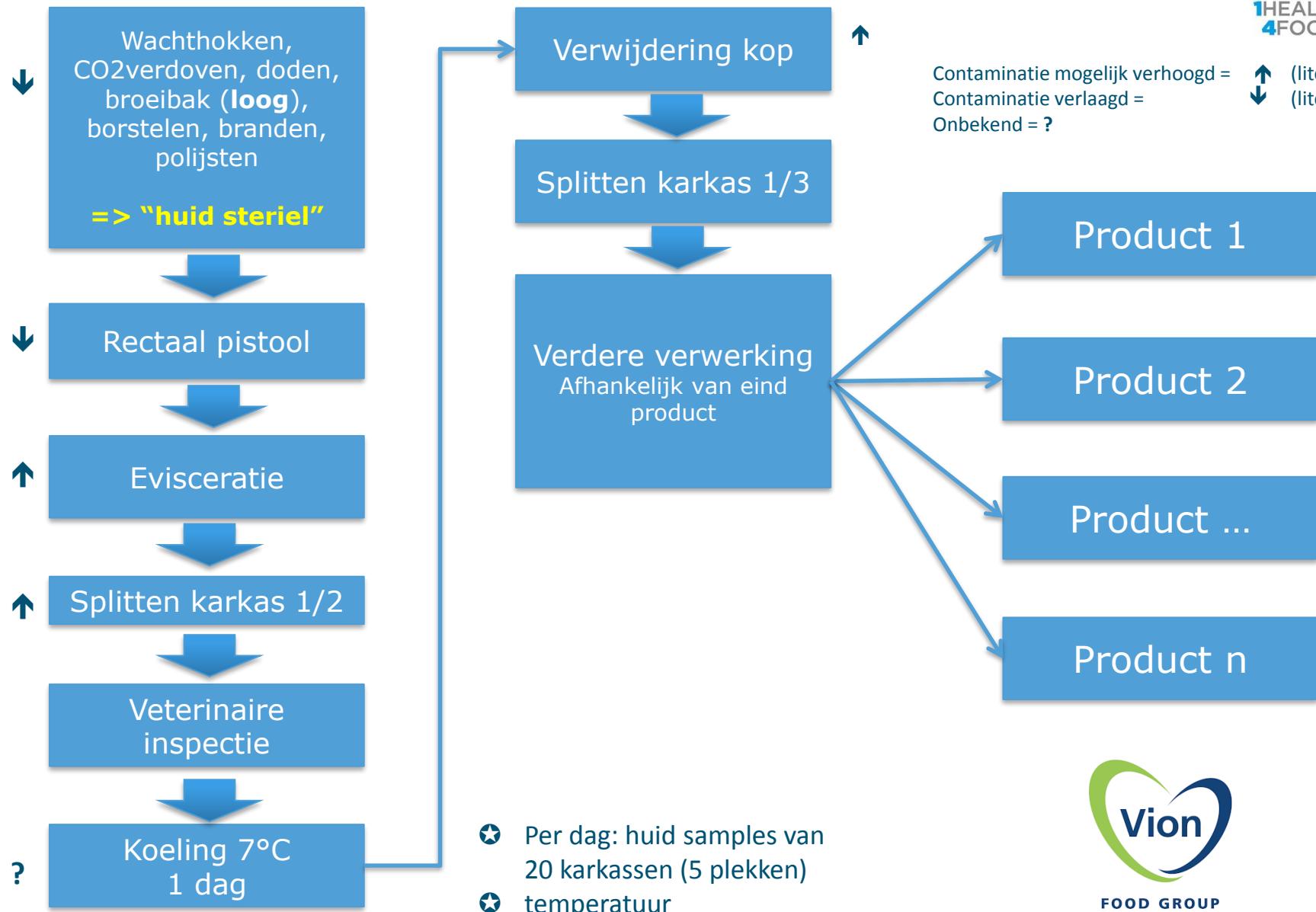


# Sequences to interpretation





## Overview meat production line Vion Boxtel



- ★ Per dag: huid samples van 20 karkassen (5 plekken)
- ★ temperatuur



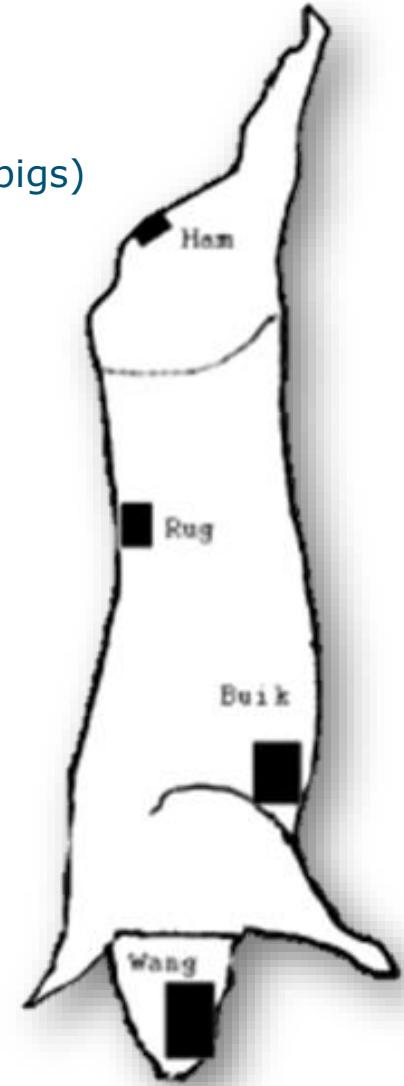
↑ (literatuur)  
↓ (literatuur)



# Sampling procedure for carcasses



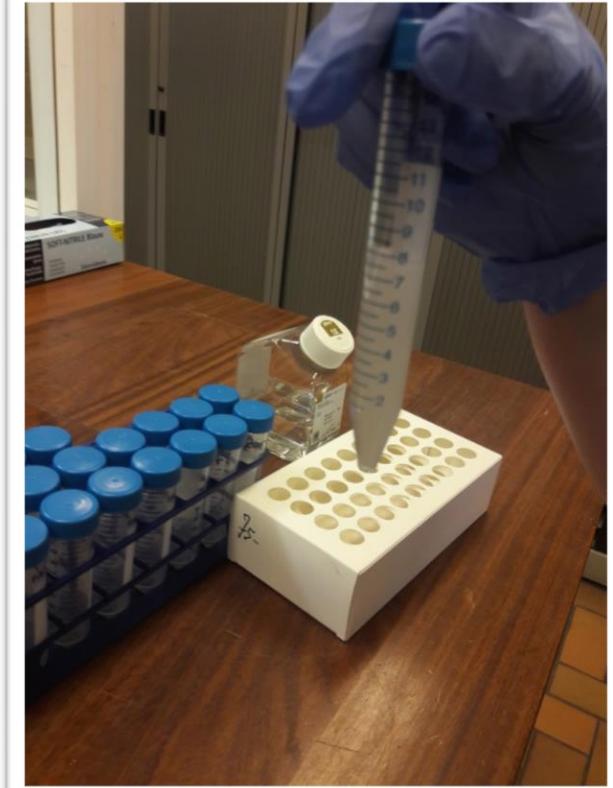
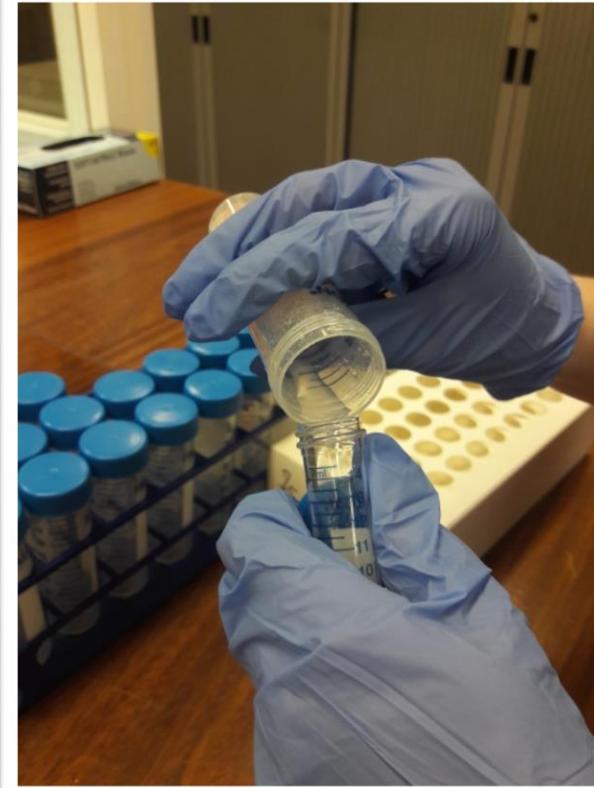
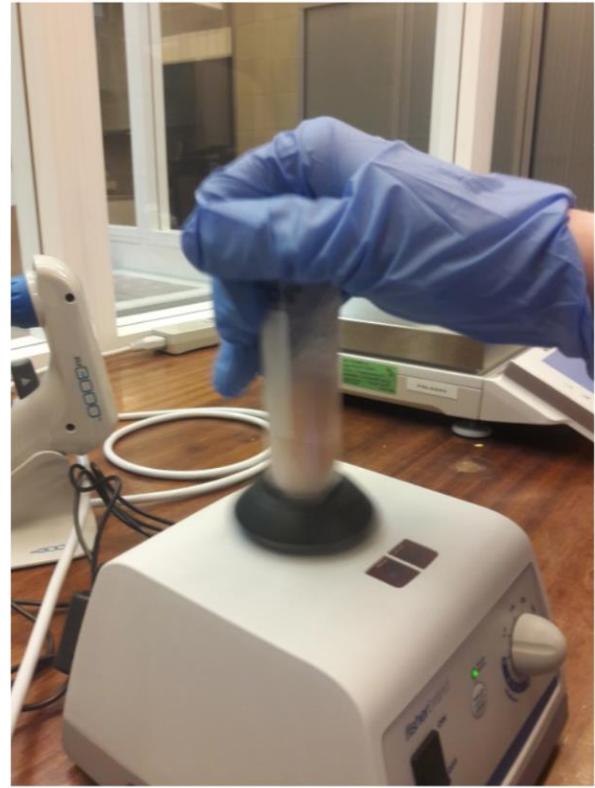
- Uniform procedure on all sites
- Trained staff
- Daily *at random* sampling ~20 carcasses in Boxtel (1 / 1,000 slaughtered pigs)  
Oct 2017 – Oct 2018: ~4,100 samples collected (sampling 4 dy/week)
- Routine excision sampling:  
4× 5 cm<sup>2</sup> per carcass, for TVC and enterobacteriaceae
- Five carcasses selected for Salmonella pool: additional excision samples



# Skin patch sampling



# Skin sample prep

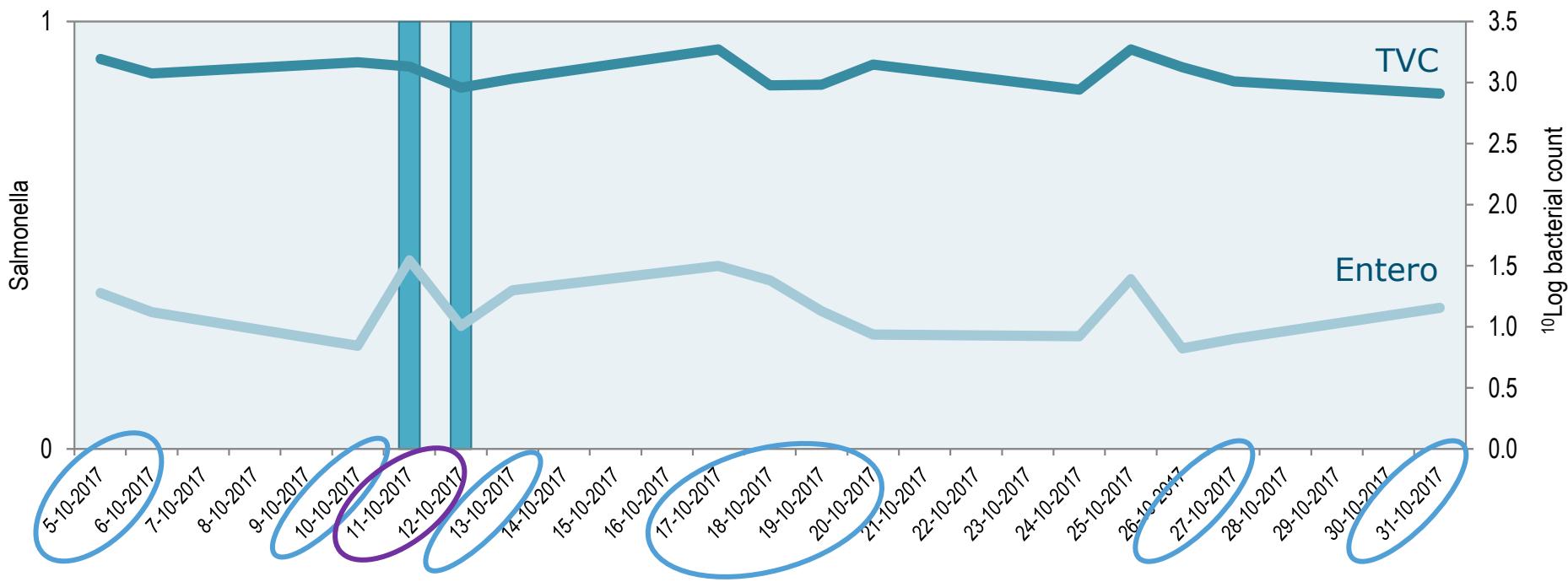


# Use case Salmonella

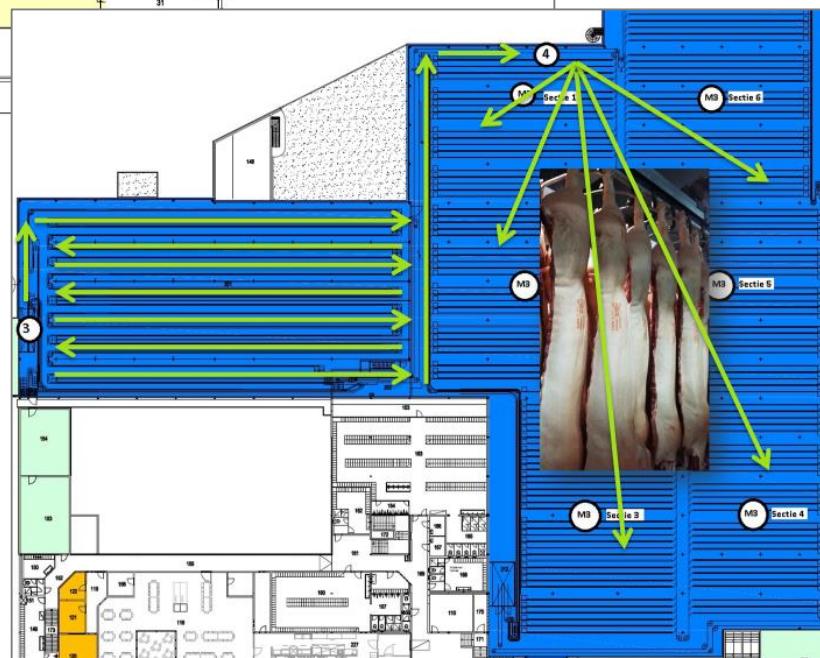
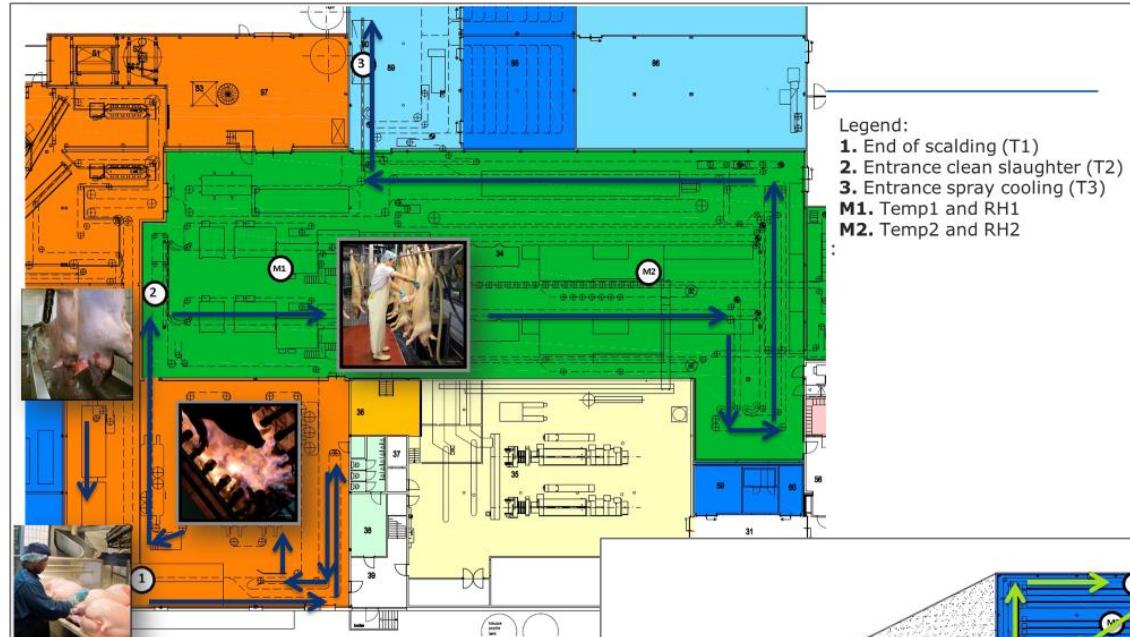
## Microbiological results



- Results obtained from 15 salmonella pools (1 month)
- 2 Salmonella positive pools detected



# Temperature & Relative humidity



# Microbiome analysis



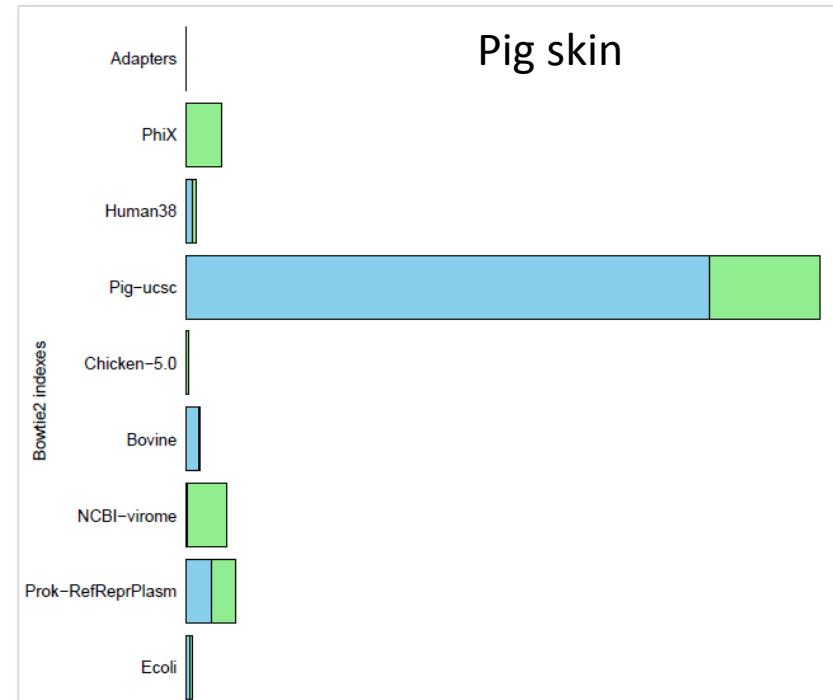
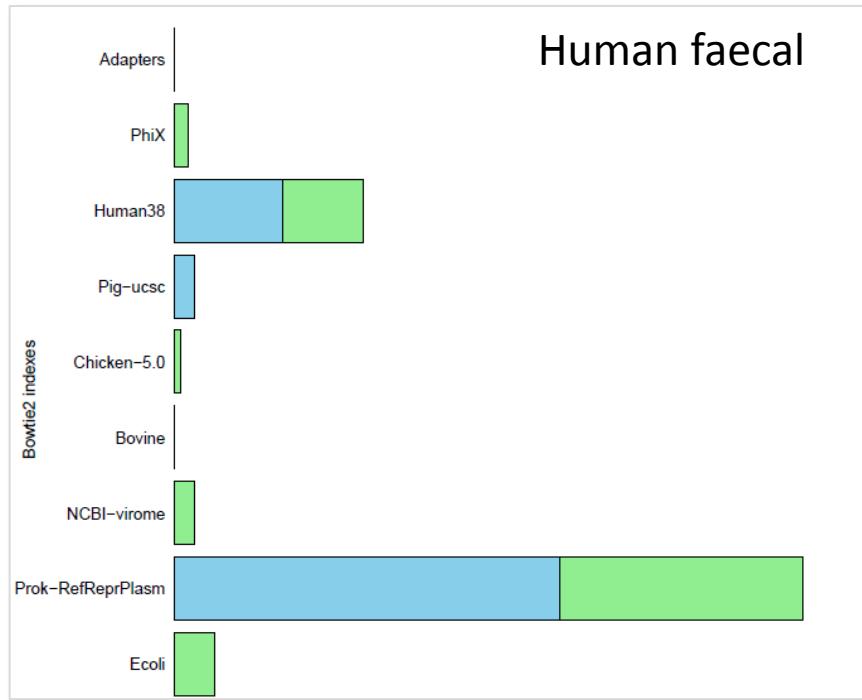
- Conditions for sample storage/prep
  - Sample suitable?
  - DNA extraction
  - Bacteriome vs full microbiome
    - 16S-barcoding vs shotgun seq (full metagenome)
- Future
  - Dead/alive microbiome
    - 16S-barcoding sample pre-treatment
    - Meta-transcriptomics

# Bigger/more = better?

## Full metagenome microbiome analysis method?



- Mapping 80M sequences against several species databases --> counts

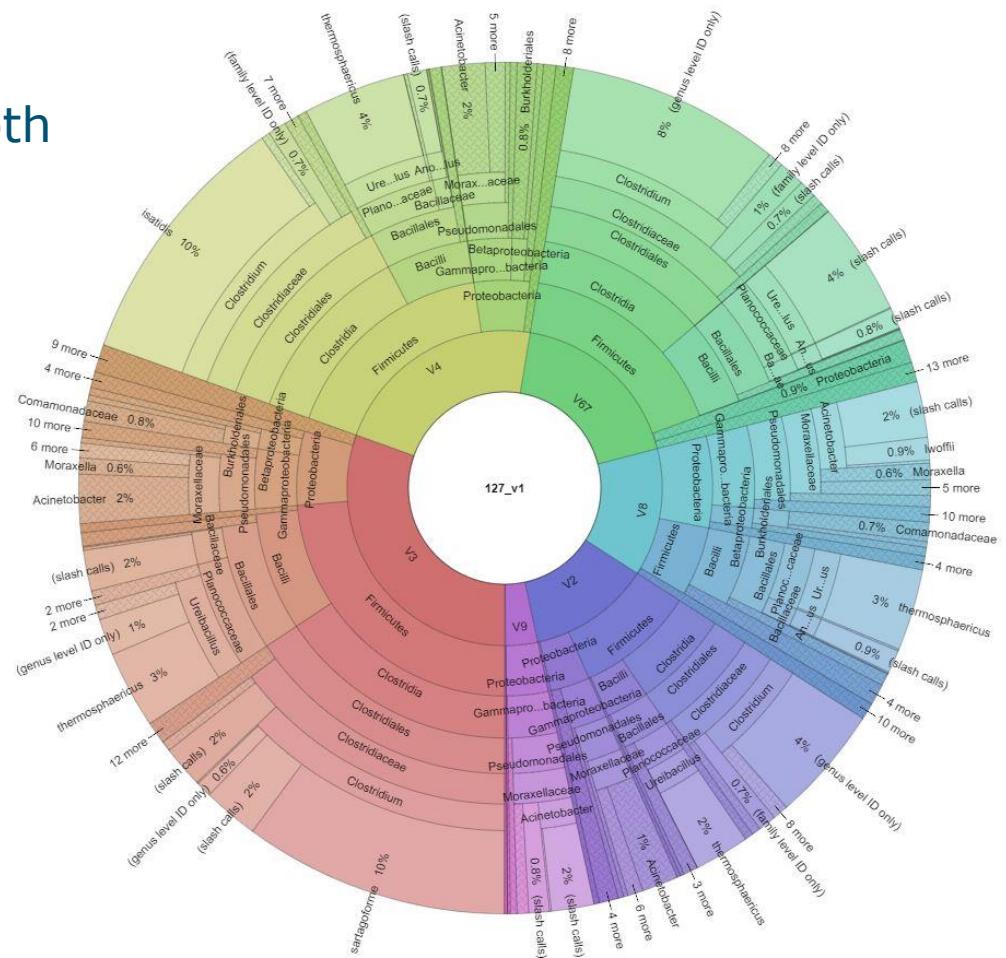


# 16S-barcoding



- Ion-torrent system (~240-400 bases/read)
- Multiple 16S rRNA gene variable regions (v2, v3, v4, v6-7, v8, v9)
- Bacteriome ~ 1-2M seq depth

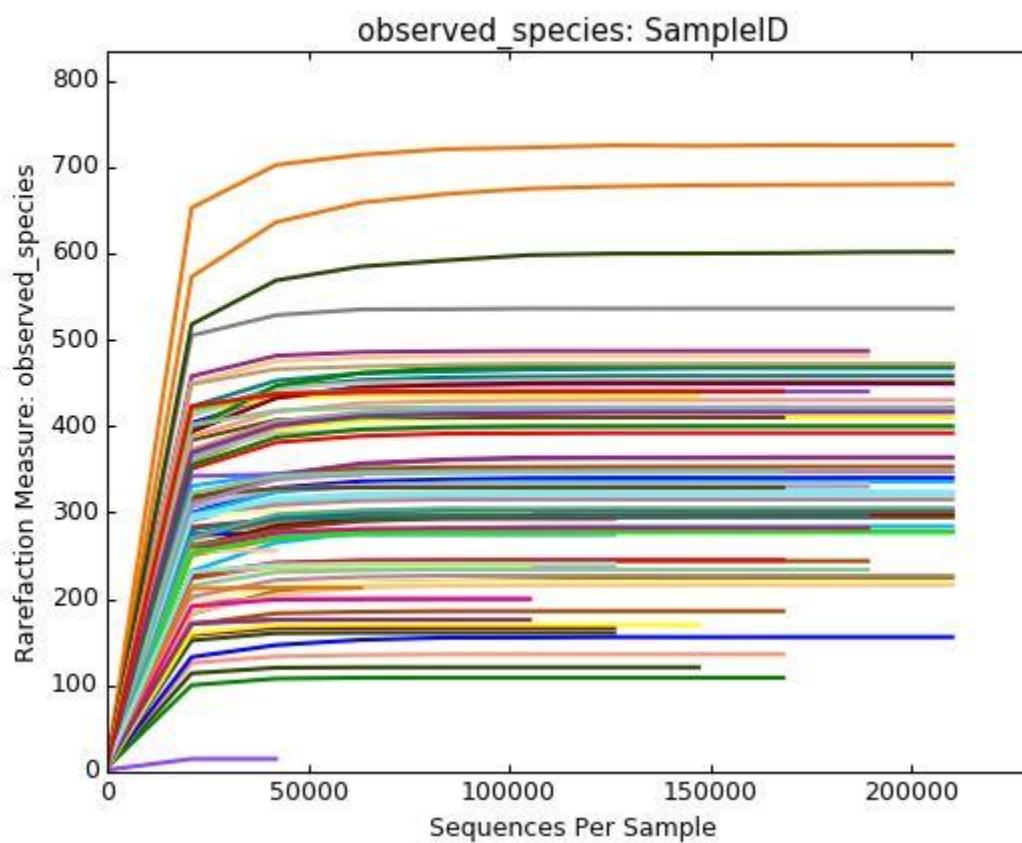
**Thermo  
Fisher**  
SCIENTIFIC



# Detected taxonomic units



Select a Metric:  Select a Category:

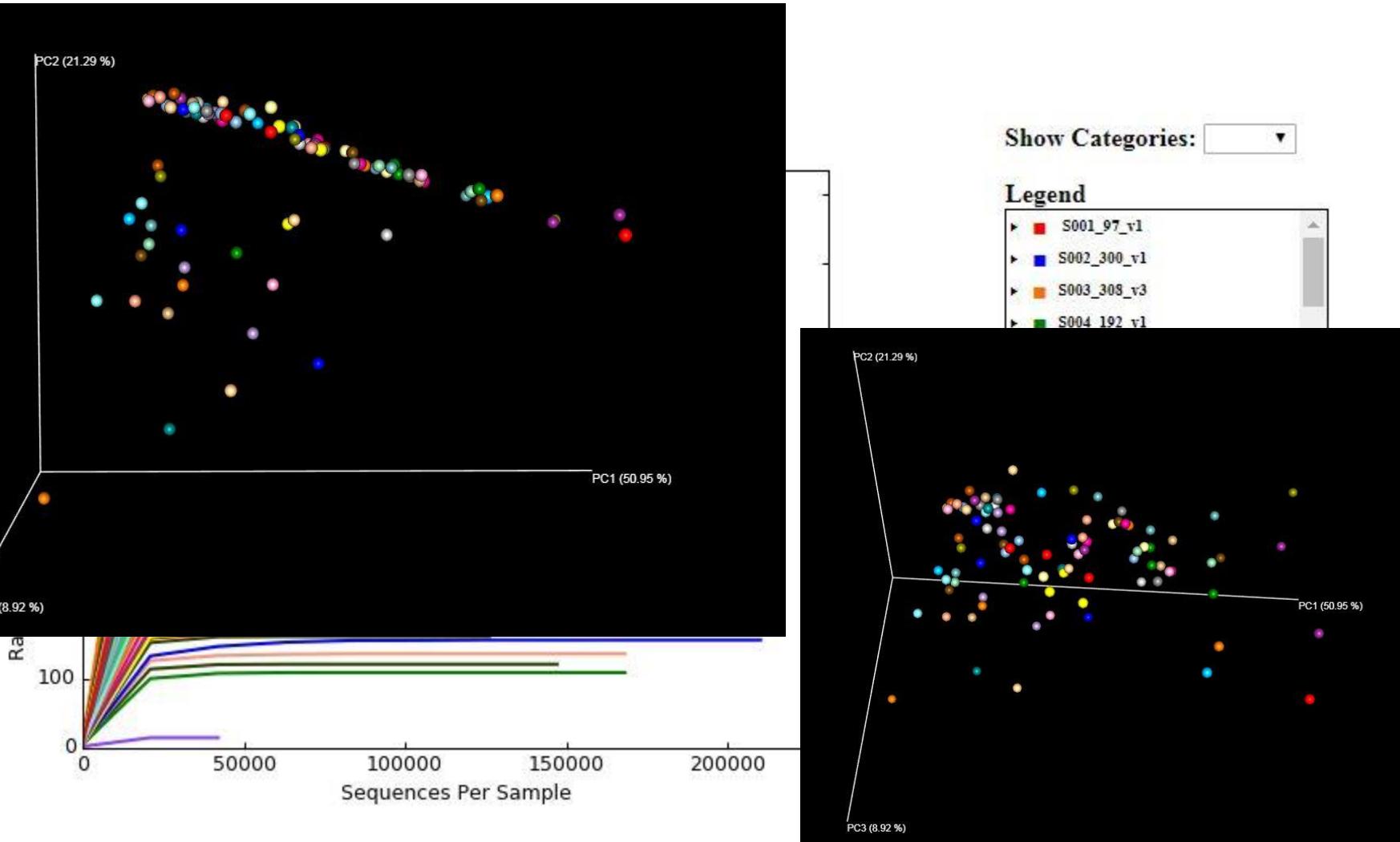


Show Categories:

## Legend

- S001\_97\_v1
- S002\_300\_v1
- S003\_308\_v3
- S004\_192\_v1
- S005\_111\_v3
- S006\_49\_v1
- S007\_80\_v1
- S009\_130\_v2
- S010\_173\_v1
- S011\_35\_v1
- S012\_148\_v1
- S013\_58\_v1
- S014\_196\_v1
- S015\_177\_v1
- S016\_310\_v1

# Detected taxonomic units

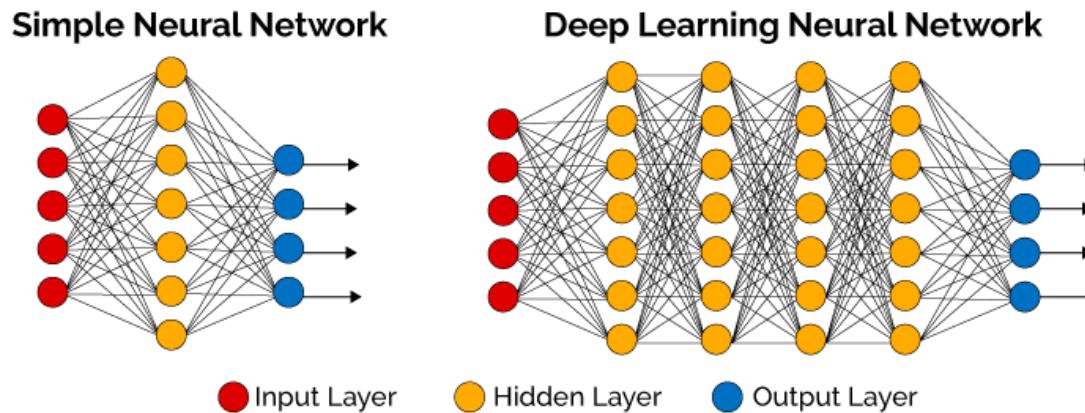


# Probably deep-learning needed

IBM – Watson :: exploratory studies



- Find non-obvious correlations --> early signatures
- Integrate other (meta) data
  - TVC, enterobacteriaceae, Salmonella count, ...
  - 16S data
  - Sensor data (temperature/humidity)

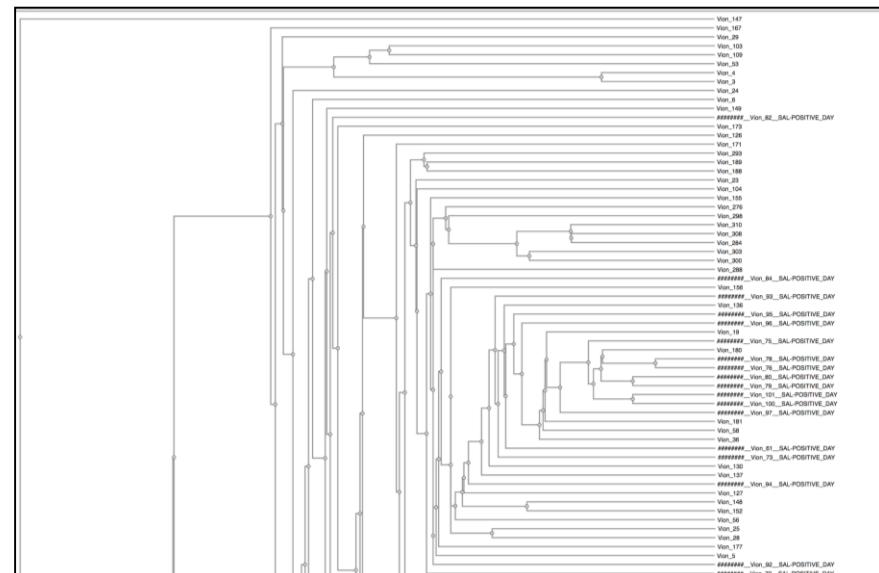
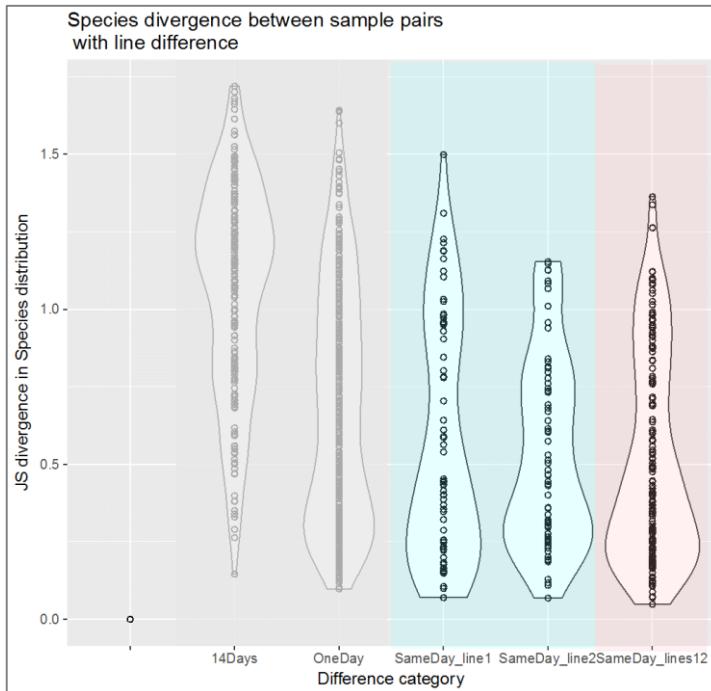


# Preliminary results

On a subset of analysed samples (first month)



- In rank-based correlations a few taxa informative (@genus)
- Microbiomes of two processing lines not different
- Reference-free clustering:  
No correlations yet



# Future

*Just started...*



- Pooling strategies to increase data coverage
- More data --> complete year
  - Integrate sensor data
  - Deep analysis for early signatures
- If early signatures exist --> biomarkers  
--> fast and cheap but still sensitive
- Intervention modelling when signatures are discovered

# Thank you for your attention



## WBVR

Alex Bossers  
Cynthia Ho  
Albert de Boer

## WU-FM

Marcel Zwietering  
Tjakko Abée



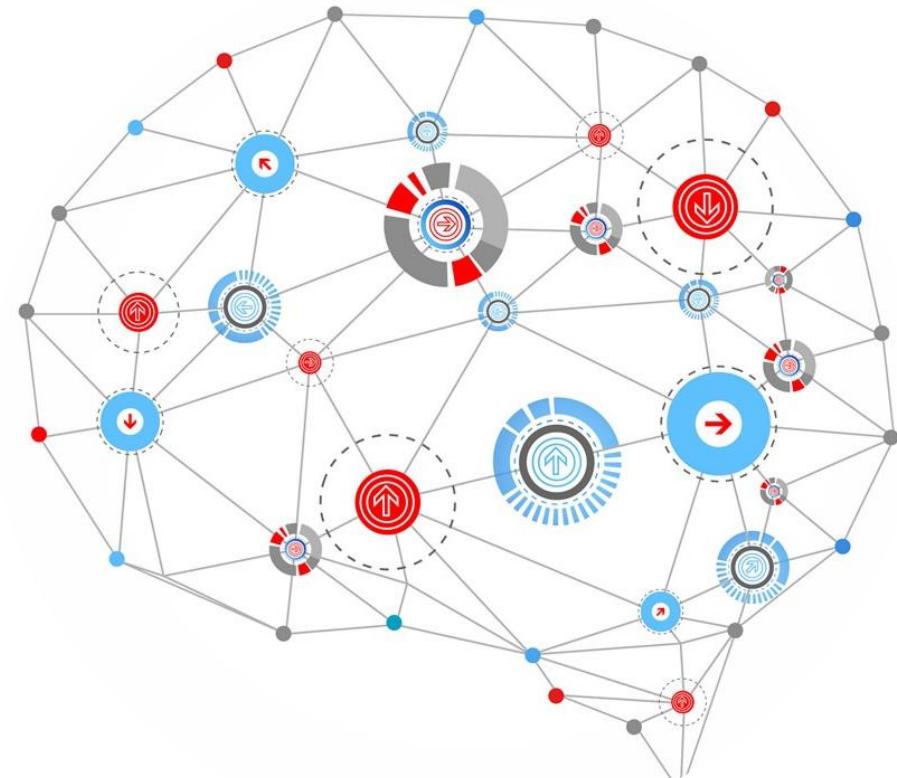
Bert Urlings  
Martijn Bouwknegt



Patrik Buholzer  
Livio Baselgia



David Chambliss  
Nimrod Megiddo  
Ban Kawas

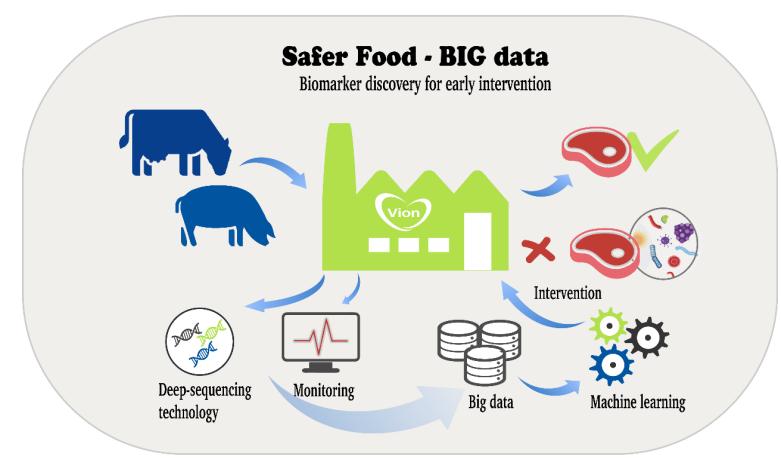


# Stelling 1



■ BIG-data toepassingen zijn operationeel in het food-safety domein binnen:

- <5 jaar
- 5-10 jaar
- >10 jaar
- Nooit



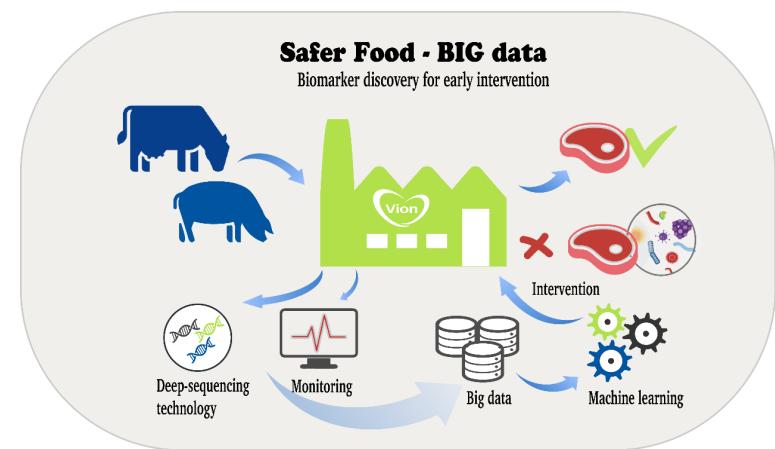
# Stelling 2



## ■ Klassieke microbiologie rules!

*En zal altijd nodig blijven.*

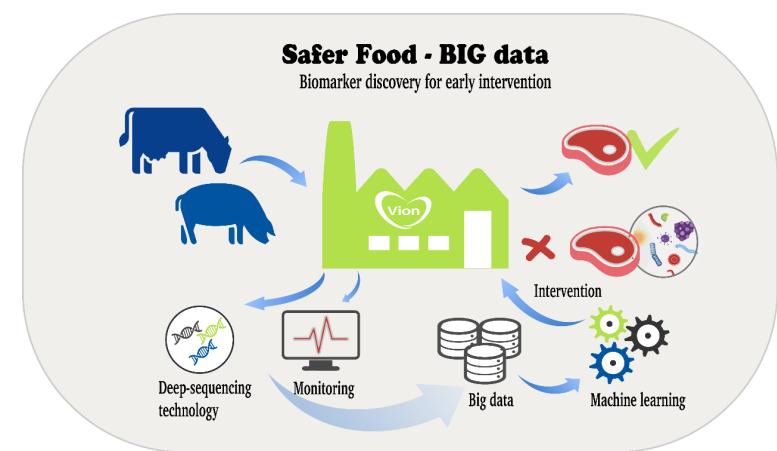
- Eens
- Niet eens
- Weet niet



# Stelling 3



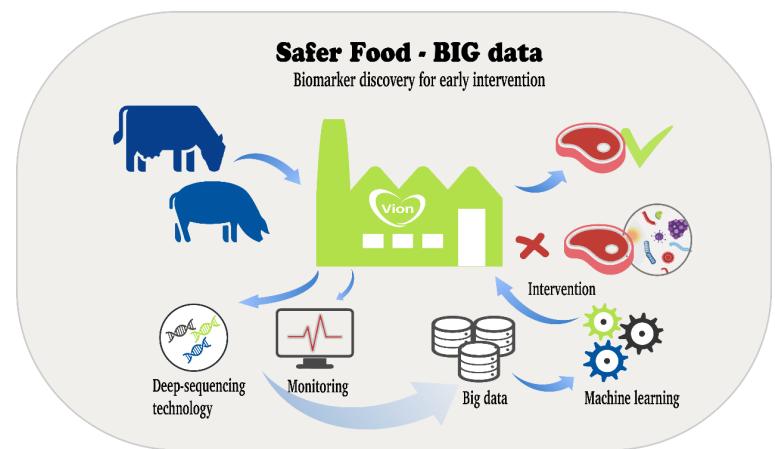
- Next generation sequencing in food safety is een *pandorra's box*
  - Eens
  - Oneens
  - Geen mening



# Stelling 4

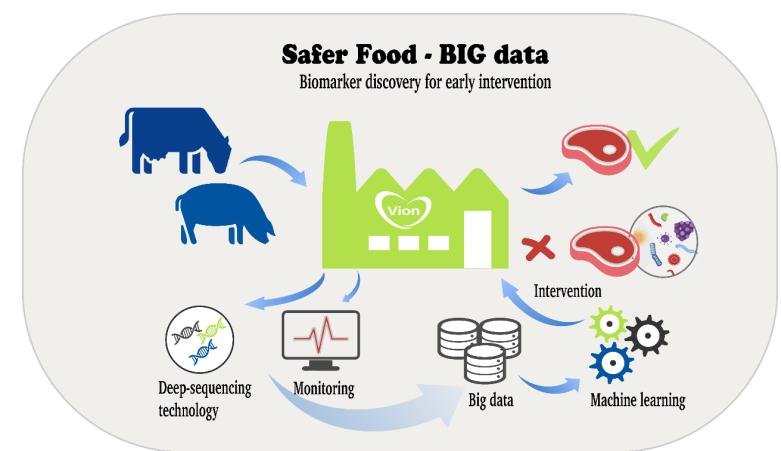


- The BIGGER the data, the BETTER
  - Eens
  - Oneens
  - Geïntegreerde data is belangrijker



# Stelling 5

- Correlaties via deep-learning applicaties zijn niet voldoende, causaties zijn nodig
  - Eens
  - Oneens





# What else?

estNreadClade	#clade_name					
85361	k_Viruses					
112043	k_Bacteria					
658	k_Eukaryota					
85361	k_Viruses	p_Viruses_noname				
102653	k_Bacteria	p_Actinobacteria				
9839	k_Bacteria	p_Proteobacteria				
4473	k_Bacteria	p_Firmicutes				
405	k_Eukaryota	p_Apicomplexa				
9449	k_Viruses	p_Viruses_noname	c_Viruses_noname	o_Viruses_noname	f_Retroviridae	g_Gammaretrovirus
810	k_Viruses	p_Viruses_noname	c_Viruses_noname	o_Viruses_noname	f_Partitiviridae	g_Alphacryptovirus
77633	k_Bacteria	p_Actinobacteria	c_Actinobacteria	o_Actinomycetales	f_Propionibacteriaceae	g_Propionibacterium
215	k_Viruses	p_Viruses_noname	c_Viruses_noname	o_Viruses_noname	f_Potyviridae	g_Potyvirus
371	k_Viruses	p_Viruses_noname	c_Viruses_noname	o_Viruses_noname	f_Viruses_noname	g_Viruses_noname
8878	k_Bacteria	p_Proteobacteria	c_Gammaproteobacteria	o_Pseudomonadales	f_Moraxellaceae	g_Acinetobacter
5148	k_Bacteria	p_Actinobacteria	c_Actinobacteria	o_Actinomycetales	f_Propionibacteriaceae	g_Propionibacteriaceae_unclassified
3697	k_Bacteria	p_Firmicutes	c_Bacilli	o_Bacillales	f_Staphylococcaceae	g_Staphylococcus
2970	k_Bacteria	p_Proteobacteria	c_Gammaproteobacteria	o_Pseudomonadales	f_Pseudomonadaceae	g_Pseudomonas
13	k_Viruses	p_Viruses_noname	c_Viruses_noname	o_Caudovirales	f_Podoviridae	g_P22likevirus
598	k_Bacteria	p_Actinobacteria	c_Actinobacteria	o_Actinomycetales	f_Corynebacteriaceae	g_Corynebacterium
2	k_Viruses	p_Viruses_noname	c_Viruses_noname	o_Viruses_noname	f_Flaviviridae	g_Flavivirus
455	k_Bacteria	p_Firmicutes	c_Bacilli	o_Bacillales	f_Planoctococcaceae	g_Ureibacillus
22	k_Viruses	p_Viruses_noname	c_Viruses_noname	o_Viruses_noname	f_Baculoviridae	g_Alphabaculovirus
3	k_Viruses	p_Viruses_noname	c_Viruses_noname	o_Caudovirales	f_Siphoviridae	g_C2likevirus
723	k_Bacteria	p_Proteobacteria	c_Gammaproteobacteria	o_Enterobacterales	f_Enterobacteriaceae	g_Salmonella
331	k_Bacteria	p_Actinobacteria	c_Actinobacteria	o_Actinomycetales	f_Micrococcaceae	g_Rothia
258	k_Bacteria	p_Firmicutes	c_Bacilli	o_Lactobacillales	f_Streptococcaceae	g_Streptococcus
293	k_Bacteria	p_Proteobacteria	c_Gammaproteobacteria	o_Pseudomonadales	f_Moraxellaceae	g_Enhydrobacter
308	k_Bacteria	p_Actinobacteria	c_Actinobacteria	o_Actinomycetales	f_Corynebacteriaceae	g_Turicella
441	k_Eukaryota	p_Apicomplexa	c_Aconoidasida	o_Haemosporida	f_Haemosporida_noname	g_Plasmodium
51	k_Bacteria	p_Proteobacteria	c_Gammaproteobacteria	o_Aeromonadales	f_Aeromonadaceae	g_Aeromonas

# Biofilm: trouble ~ informer

